

Chapter 2

Role of Enzymes in Maintaining Soil Health

Shonkor Kumar Das and Ajit Varma

2.1 Introduction

Enzymes are the vital activators in life processes, likewise in the soil they are known to play a substantial role in maintaining soil health and its environment. The enzymatic activity in the soil is mainly of microbial origin, being derived from intracellular, cell-associated or free enzymes. A unique balance of chemical, physical, and biological (including microbial especially enzyme activities) components contribute to maintaining soil health. Evaluation of soil health therefore requires indicators of all these components. Healthy soils are essential for the integrity of terrestrial ecosystems to remain intact or to recover from disturbances, such as drought, climate change, pest infestation, pollution, and human exploitation including agriculture (Ellert et al. 1997). Deterioration of soil, and thereby soil health, is of concern for human, animal, and plant health because air, groundwater, and surface water consumed by humans, can be adversely affected by mismanaged and contaminated soil (Singer and Ewing 2000). As soil is the part of the terrestrial environment and supports all terrestrial life forms, protection of soil is therefore of high priority and a thorough understanding of soil enzymes activities is a critical factor in assuring that soil remains healthy. A better understanding of the role of this soil enzymes activity in maintaining the soil health will potentially provide a unique opportunity for an integrated biological assessment of soils due to their crucial role in several soil biological activities, their ease of measurement, and their rapid response to changes in soil management. Although there have been extensive studies on soil enzymes, little has been reported on their roles in maintaining soil health. Thus, it is authoritative to

S.K. Das (✉)

Department of Applied Chemistry and Biotechnology, Graduate School of Engineering, University of Fukui, 3-9-1, Bunkyo, Fukui 910-8507, Japan
e-mail: skdas76@yahoo.com

A. Varma

Amity Institute of Microbial Technology, Amity University, Sector 125, Noida 201303, Uttar Pradesh, India

understand the roles of these enzymes' activity and their efficiency to maintain soil health for future betterment of soil research and soil biology.

2.2 Soil Enzymes

Soil enzymes are a group of enzymes whose usual inhabitants are the soil and are continuously playing an important role in maintaining soil ecology, physical and chemical properties, fertility, and soil health. These enzymes play key biochemical functions in the overall process of organic matter decomposition in the soil system (Sinsabaugh et al. 1991). They are important in catalyzing several vital reactions necessary for the life processes of micro-organisms in soils and the stabilization of soil structure, the decomposition of organic wastes, organic matter formation, and nutrient cycling, hence playing an important role in agriculture (Dick et al. 1994; Dick 1997).

All soils contain a group of enzymes that determine soil metabolic processes (McLaren 1975) which, in turn, depend on its physical, chemical, microbiological, and biochemical properties. The enzyme levels in soil systems vary in amounts primarily due to the fact that each soil type has different amounts of organic matter content, composition, and activity of its living organisms and intensity of biological processes. In practice, the biochemical reactions are brought about largely through the catalytic contribution of enzymes and variable substrates that serve as energy sources for microorganisms (Kiss et al. 1978). These enzymes may include amylase, arylsulphatases, β -glucosidase, cellulose, chitinase, dehydrogenase, phosphatase, protease, and urease released from plants (Miwa et al. 1937), animals (Kanfer et al. 1974), organic compounds, and microorganisms (James et al. 1991; Richmond 1991; Shawale and Sadana 1981) and soils (Gupta et al. 1993; Ganeshamurthy et al. 1995).

2.2.1 *Kind of Soil Enzymes*

2.2.1.1 Constitutive

Always present in nearly constant amounts in a cell (not affected by addition of any particular substrate – genes always expressed).

(Pyrophosphatase)

2.2.1.2 Inducible

Present only in trace amounts or not at all, but quickly increases in concentration when its substrate is present.

(Amidase)

Both types of enzymes are present in the soil.

2.2.2 Origin and State of Soil Enzymes

Although the general origins of soil enzymes are (a) microorganisms-living and dead, (b) plant roots and plant residues and, (c) soil animals; the state of soil enzymes in the soil is different as below

State-1: Role of clays

Most activity associated with clays

Increased resistance to proteolysis and microbial attack

Increases the temperature of inactivation

State-2: Role of organic matter

Humus material provides stability to soil nitrogen compounds

Enzymes attached to insoluble organic matrices exhibit pH and temperature changes

Inability to purify soil enzymes free of soil organic matters (bound to organic matter)

State-3: Role of clay–organic matter complexes

Lignin + bentonite (clay) protect enzymes against proteolytic attack, but not bentonite alone

Enzymes are bound to organic matter which is then bound to clay

2.2.3 Importance of Soil Enzymes

Release of nutrients into the soil by means of organic matter degradation

Identification of soils

Identification of microbial activity

Importance of soil enzymes as sensitive indicators of ecological change

2.2.4 Application of Soil Enzymes

Correlation with soil fertility

Correlation with microbial activity

Correlation with biochemical cycling of various elements in soil (C, N, S)

Degree of pollution (heavy metals, SO₄)

To assess the successional stages of an ecosystem

Forensic purposes

Rapid degradation of pesticides

Disease studies

Enzyme activity in soil fluctuates with environment.

2.3 Soil Health

2.3.1 Definition

The definition of soil health must be broad enough to encompass the many functions of soil, e.g., environmental filter, plant growth, and water regulation (Doran and Safley 1997). Definitions of air and water quality standards have existed for a long time, while a similar definition does not exist for soil. A definition of soil health based on this concept would encompass only a small fraction of the many roles soil play (Singh et al. 1999). Soil health is the net result of on-going conservation and degradation processes, depending highly on the biological component of the soil ecosystem, and influences plant health, environmental health, food safety, and quality (Halvorson et al. 1997; Parr et al. 1992).

Several definitions of soil health have been proposed during the last decades. Historically, the term soil quality described the status of soil as related to agricultural productivity or fertility (Singer et al. 1999). In the 1990s, it was proposed that soil quality was not limited to soil productivity but instead expanded to encompass interactions with the surrounding environment, including the implications for human and animal health. In this regard, several examples of definitions of soil quality have been suggested (Doran and Parkin 1994). In the mid-1990s, the term *soil health* was introduced. For example, a program to assess and monitor soil health in Canada used the terms quality and health synonymously to describe the ability of soil to support crop growth without becoming degraded or otherwise harming the environment (Acton and Gregorich 1995). Others broadened the definition of soil health to capture the ecological attributes of soil, and went beyond its capacity to simply produce particular crops. These attributes are chiefly associated with biodiversity, food web structure, and functional measures (Pankhurst et al. 1997).

Several numbers have been recognized for soil health which are as follows:

1. The continued capacity of soil to function as a vital living system, within the ecosystem and land-use boundaries, to sustain biological productivity, promote the quality of air and water environments, and maintain plant, animal, and human health (Doran and Safley 1997).
2. Soil health is an assessment of ability of a soil to meet its range of ecosystem functions as appropriate to its environment.
3. Soil health can also be defined as the continued capacity of a specific kind of soil to function as a vital living system, within natural or managed ecosystem boundaries, to sustain plant and animal productivity, to maintain or enhance the quality of air and water environments, and to support human health and habitation.

2.3.2 Aspects of Soil Health

The term soil health is used to assess the ability of a soil to

Sustain plant and animal productivity and diversity

Maintain or enhance water and air quality

Support human health and habitation

The underlying principle in the use of the term “soil health” is that soil is not just a growing medium; rather, it is a living, dynamic and ever-so-subtly changing environment. We can use the human health analogy and categorize a healthy soil as one

In a state of composite well-being in terms of biological, chemical, and physical properties

Not diseased or infirmed (i.e., not degraded, nor degrading), nor causing negative off-site impacts

With each of its qualities cooperatively functioning such that the soil reaches its full potential and resists degradation

Providing a full range of functions (especially nutrient, carbon, and water cycling), and in such a way that it maintains this capacity into the future.

2.3.3 Interpretation of Soil Health

Different soils will have different benchmarks of health depending on the “inherited” qualities, and on the geographic circumstance of the soil. The generic aspects defining a healthy soil can be considered as follows

“Productive” options are broad

Life diversity is broad

Absorbency, storing, recycling, and processing is high in relation to limits set by climate

Water runoff quality is of high standard

Low entropy

No damage to or loss of the fundamental components

This translates to

A comprehensive cover of vegetation

Carbon levels relatively close to the limits set by soil type and climate

Little leakage of nutrients from the ecosystem

Biological productivity relatively close to the limits set by the soil environment and climate

Only geological rates of erosion

No accumulation of contaminants

The ecosystem does not rely excessively on inputs of fossil energy

An unhealthy soil thus is the simple converse of the above.

2.3.4 Pressures on Soil Health Towards Impacts

The flow chart given below is a simple description of soil health factors and their impacts (Fig. 2.1).

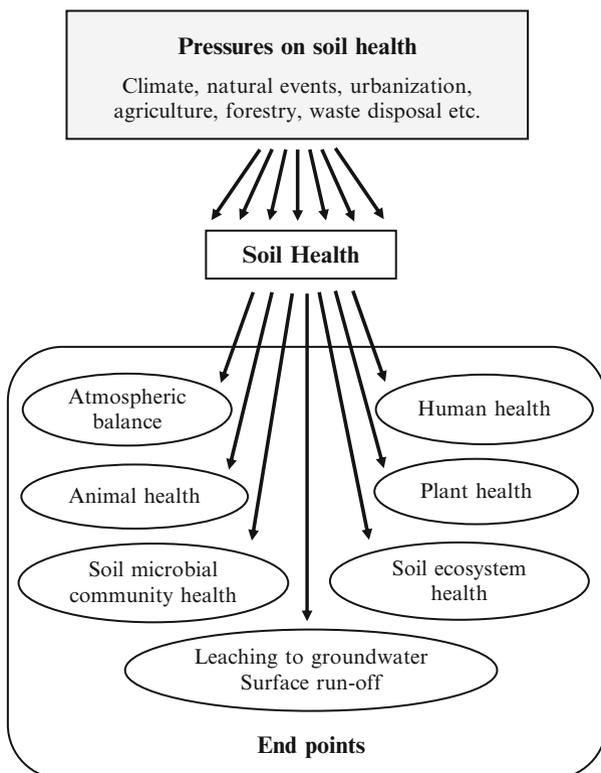


Fig. 2.1 Policy relevant end points of soil health monitoring

2.4 Indicators of Soil Health

2.4.1 *Microorganism as Indicators of Soil Health*

The biological activity in soil is largely concentrated in the topsoil, the depth of which may vary from a few to 30 cm. In the topsoil, the biological components occupy a tiny fraction (<0.5%) of the total soil volume and make up less than 10% of the total organic matter in the soil. These biological components consist mainly of soil organisms, especially microorganisms. Despite of their small volume in soil, microorganisms are key players in the cycling of nitrogen, sulfur, and phosphorus, and the decomposition of organic residues. Thereby they affect nutrient and carbon cycling on a global scale (Pankhurst et al. 1997). That is, the energy input into the soil ecosystems is derived from the microbial decomposition of dead plant and animal organic matter. The organic residues are, in this way, converted to biomass or mineralized to CO₂, H₂O, mineral nitrogen, phosphorus, and other nutrients (Bloem et al. 1997). Microorganisms are further associated with the transformation and degradation of waste materials and synthetic organic compounds (Torstensson et al. 1998). In addition to the effect on nutrient cycling, microorganisms also affect the physical properties of the soil. Production of extra-cellular polysaccharides and other cellular debris by microorganisms help in maintaining soil structure as well as soil health. Thereby, they also affect water holding capacity, infiltration rate, crusting, erodibility, and susceptibility to compaction (Elliott et al. 1996).

Microorganisms possess the ability to give an integrated measure of soil health, an aspect that cannot be obtained with physical/chemical measures and/or analyses of diversity of higher organisms. Microorganisms respond quickly to changes; hence they rapidly adapt to environmental conditions, and thus they can be used for soil health assessment, and changes in microbial populations and activities may therefore function as an excellent indicator of change in soil health (Kennedy and Papendick 1995; Pankhurst et al. 1995).

Microorganisms also respond quickly to environmental stress compared to higher organisms, as they have intimate relations with their surroundings due to their high surface to volume ratio. In some instances, changes in microbial populations or activity can precede detectable changes in the soil's physical and chemical properties, thereby providing an early sign of soil improvement or an early warning of soil degradation (Pankhurst et al. 1995). The impact of some chemicals on soil health is dependent on microbial activities. For example, the concentration of heavy metals in soil will not change over small time periods, but their bioavailability may. In this way, soil enzymes are acting as important indicators of soil.

2.4.2 *Soil Enzymes as Indicators of Soil Health*

Enzymes are the direct mediators for biological catabolism of soil organic and mineral components. Thus, these catalysts provide a meaningful assessment of

Table 2.1 Soil enzymes as indicators of soil health

Soil enzyme	Enzyme reaction	Indicator of microbial activity
Dehydrogenase	Electron transport system	C-cycling
β -glucosidase	Cellobiose hydrolysis	C-cycling
Cellulase	Cellulose hydrolysis	C-cycling
Phenol oxidase	Lignin hydrolysis	C-cycling
Urease	Urea hydrolysis	N-cycling
Amidase	N-mineralization	N-cycling
Phosphatase	Release of PO_4^-	P-cycling
Arylsulphatase	Release of SO_4^-	S-cycling
Soil enzymes	Hydrolysis	General organic matter degradative enzyme activities

reaction rates for important soil processes. Soil enzyme activities (1) are often closely related to soil organic matter, soil physical properties and microbial activity or biomass, (2) changes much sooner than other parameters, thus providing early indications of changes in soil health, and (3) involve simple procedures (Dick et al. 1996). In addition, soil enzyme activities can be used as measures of microbial activity, soil productivity, and inhibiting effects of pollutants (Tate 1995). Easy, well-documented assays are available for a large number of soil enzyme activities (Dick et al. 1996; Tabatabai 1994a, b). These include dehydrogenase, glucosidases, urease, amidases, phosphatases, arylsulphatase, cellulases, and phenol oxidases as shown in Table 2.1.

2.5 Potential Roles of Soil Enzymes in Maintaining Soil Health

A number of soil enzymes and their respective roles in maintaining soil health are stated below

2.5.1 Amylase

The starch hydrolyzing enzyme amylase (Ross 1976) is known to be constituted by α -amylase and β -amylase (King 1967; Thoma et al. 1971). The α -amylases are synthesized by plants, animals, and microorganisms, whereas, β -amylase is synthesized mainly by plants (Pazur 1965; Thoma et al. 1971). This enzyme is widely distributed in plants and soils so it plays a significant role in the breakdown of starch, which converts starch like substrates to glucose and/or oligosaccharides and β -amylase, which converts starch to maltose (Thoma et al. 1971). Studies have, however, indicated that the roles and activities of α -amylase and β -amylase enzymes may be influenced by different factors ranging from cultural practices, type of vegetation, environment and soil types (Pancholy and Rice 1973; Ross 1975). For example, plants may influence the amylase enzyme activities of soil by

directly supplying enzymes from their residues or excreted compounds, or indirectly providing substrates for the synthetic activities of microorganisms. Greater understanding is required of the significance of these enzymes in the soil, and to enable proper management techniques to be devised to maximize the benefits that may be derived from such enzymes.

2.5.2 *Arylsulphatases*

This is due to the fact that certain proportions of sulphur in different soil profiles are bound into organic compounds and are indirectly available to plants. Arylsulphatases are typically widespread in nature (Dodgson et al. 1982) as well as in soils (Gupta et al. 1993; Ganeshamurthy et al. 1995). They are responsible for the hydrolysis of sulphate esters in the soil (Kertesz and Mirleau 2004) and are secreted by bacteria into the external environment as a response to sulphur limitation (McGill and Colle 1981). Its occurrence in different soil systems is often correlated with microbial biomass and rate of S immobilization (Klose and Tabatabai 1999; Vong et al. 2003). This enzyme has a role in the hydrolysis of aromatic sulphate esters ($R-O-SO_3^-$) to phenols ($R-OH$) and sulfate, or sulfate sulfur (SO_4^{2-} or SO_4-S) (Tabatabai 1994a, b).

Soil is affected by various environmental factors (Burns 1982) such as heavy metal pollution (Tyler 1981); pH changes in the soil solution (Acosta-Martínez and Tabatabai 2000); organic matter content and its type (Sarathchandra and Perrott 1981); such as absorption to particles surfaces in soils, and the activity persistence of extra cellular arylsulfatases in the soil. Considering the importance of S in plant nutrition, a better understanding of the role(s) of arylsulfatases in S mobilization in agricultural soils is critical. So far, very little is known about specific microbial genera or species that play an important role in the soil organosulphur circle (Kertesz and Mirleau 2004) in which arylsulphatases is the key enzyme.

2.5.3 *β -Glucosidase*

Glucosidase is a common and predominant enzyme in soils (Eivazi and Tabatabai 1988; Tabatabai 1994a, b). It is named according to the type of bond that it hydrolyses. This enzyme plays an important role in soils because it is involved in catalyzing the hydrolysis and biodegradation of various β -glucosidase present in plant debris decomposing in the ecosystem (Ajwa and Tabatabai 1994; Martinez and Tabatabai 1997). Its final product is glucose, an important C energy source of life to microbes in the soil (Esen 1993). β -glucosidase is characteristically useful as a soil quality indicator, and may give a reflection of past biological activity, the capacity of soil to stabilize the soil organic matter, and can be used to detect management effect on soils (Bandick and Dick 1999; Ndiaye et al. 2000). This has greatly facilitated its

adoption for soil quality testing (Bandick and Dick 1999). Some of the aglycons are known to be the precursors of the toxic substances, which cause soil sickness where plants are grown as monocrops (Patrick 1955; Borner 1958).

β -Glucosidase enzyme is very sensitive to changes in pH, and soil management practices (Acosta-Martínez and Tabatabai 2000; Madejón et al. 2001). Acosta-Martínez and Tabatabai 2000 reported β -glucosidase as sensitive to pH changes. This property can be used as a good biochemical indicator for measuring ecological changes resulting from soil acidification in situations involving activities of this enzyme. Consequently, more understanding of the β -glucosidase enzyme activities and factors influencing them in the ecosystem may contribute significantly to soil health studies.

2.5.4 Cellulases

Cellulose is the most abundant organic compound in the biosphere, comprising almost 50% of the biomass synthesized by photosynthetic fixation of CO₂ (Eriksson et al. 1990). Growth and survival of microorganisms important in most agricultural soils depends on the carbon source contained in the cellulose occurring in the soils (Deng and Tabatabai 1994). However, for carbon to be released as an energy source for use by the microorganisms, cellulose in plant debris has to be degraded into glucose, cellobiose and high molecular weight oligosaccharides by cellulases enzymes (White 1982). Cellulases are a group of enzymes that catalyze the degradation of cellulose, polysaccharides built up of β -1,4 linked glucose units (Deng and Tabatabai 1994). It has been reported that cellulases in soils are derived mainly from plant debris incorporated into the soil, and that a limited amount may also originate from fungi and bacteria in soils (Richmond 1991). Demonstrating the effects of increasing concentrations of fungicides on cellulases activities, Petker and Rai (1992) showed that there was a decreasing effect with fungicides captan, cosan, thiram, zinels, and sandolex. More recently, Arinze and Yubedee (2000) reported that fungicides benlate, calixin, and captan inhibited cellulase activity in *Fusarium moniliforme* isolates. Captatol inhibited cellulase activity in the sandy loam soil (Atlas et al. 1978), and chlorothalonil showed a clear reduction in cellulase activity under flooded or non-flooded conditions (Vincent and Sisler 1968). Studies have shown that activities of cellulases in agricultural soils are affected by several factors. These include temperature, soil pH, water and oxygen contents (abiotic conditions), the chemical structure of organic matter and its location in the soil profile horizon (Deng and Tabatabai 1994; Alf and Nannipieri 1995), quality of organic matter/plant debris and soil mineral elements (Sinsabaugh and Linkins 1989; Deng and Tabatabai 1994) and the trace elements from fungicides (Deng and Tabatabai 1994; Arinze and Yubedee 2000). Srinivasulu and Rangaswamy 2006 reported a significantly more stimulatory effect of cellulases in black soil than red soil. For instance, chitin in the presence of cellulose induces the synthesis of chitinase and other cell wall lytic enzymes which promote the

release of the intramural β -glucosidase into the medium. All these findings suggest that activities of cellulases can be used to give preliminary indication of some of the physical chemical properties of soil, thus, easing agricultural soil management strategies. Since cellulases enzymes play an important role in global recycling of the most abundant polymer, cellulose in nature, it would be of critical importance to understand this enzyme better so that it may be used more regularly as a predictive tool in our soil fertility programs. More information on the role of this enzyme is needed since it is affected by different factors, which may jeopardize its involvement in the decomposition of cellulolytic materials in the soil for microbial use and improved soil health in agricultural ecosystems.

2.5.5 *Chitinase*

Chitinase or chitinolytic enzymes are key enzymes responsible for the degradation and hydrolysis of chitin (poly β -1-4-(2-ncetamido-2-deoxy)-D-glucoside). They are also considered as the major structural component of many fungal cell walls that use the hyperparasitism mechanisms against pests/pathogen attack (Chet and Henis 1975; Chet 1987). These biological agents also reduce disease-producing agents by using other mechanisms such as antibiosis or competition mechanisms (Park 1960). This agriculturally important enzyme is produced or released by various organisms including plants and microorganisms (Deshpande 1986). Its presence in different forms in the ecosystem has demonstrated its effectiveness in the control of soil-borne diseases such as *Sclerotium rolfsii* and *Rhizoctonia solani* in beans and cotton, respectively (Ordentlich et al. 1988; Shapira et al. 1989). One of the mechanisms proposed involves lytic enzymes chitinase that cause the degradation of cell walls of pathogenic fungi (Ordentlich et al. 1988; Chet et al. 1990; Singh et al. 1999). As for its role in biological control of pests, moreover, due to environmental friendliness, there are so many avenues for the application of this enzyme for maintaining soil health and consequently, increase plant growth and final yields.

2.5.6 *Dehydrogenase*

The dehydrogenase enzyme activity is commonly used as an indicator of biological activity in soils (Burns 1978). This enzyme is considered to exist as an integral part of intact cells but does not accumulate extracellularly in the soil. Dehydrogenase enzyme is known to oxidize soil organic matter by transferring protons and electrons from substrates to acceptors. These processes are the part of respiration pathways of soil microorganisms and are closely related to the type of soil and soil air-water conditions (Kandeler 1996; Glinski and Stepniewski 1985). Since these processes are the part of respiration pathways of soil microorganisms, studies on the activities of dehydrogenase enzyme in the soil is very important as it may give

indications of the potential of the soil to support biochemical processes which are essential for maintaining soil fertility as well as soil health. A study by Brzezinska et al. (1998) suggested that soil water content and temperature influence dehydrogenase activity indirectly by affecting the soil redox status. After flooding the soil, the oxygen present is rapidly exhausted so that a shift of the activity from aerobic to anaerobic microorganisms takes place. Such redox transformations are closely connected with respiration activity of soil microorganisms. They may serve as indicators of the microbiological redox systems in soils and can be considered a possible measure of microbial oxidative activity (Tabatabai 1982; Trevors 1984). For instance, lack of oxygen may trigger facultative anaerobes to initiate metabolic processes involving dehydrogenase activities and the use of Fe (III) forms as terminal electron acceptors (Bromfield 1954; Galstian and Awungian 1974), a process that may affect iron availability to plants in the ecosystem (Benckiser et al. 1984). Additionally, dehydrogenase enzyme is often used as a measure of any disruption caused by pesticides, trace elements or management practices to the soil (Reddy and Faza 1989; Wilke 1991; Frank and Malkomes 1993), as well as a direct measure of soil microbial activity (Trevors 1984; Garcia and Hernández 1997). It can also indicate the type and significance of pollution in soils. For example, dehydrogenase enzyme is high in soils polluted with pulp and paper mill effluents (McCarthy et al. 1994) but low in soils polluted with fly ash (Pitchel and Hayes 1990). Similarly, higher activities of dehydrogenases have been reported at low doses of pesticides, and lower activities of the enzyme at higher doses of pesticides (Baruah and Mishra 1986).

2.5.7 Phosphatases

In soil ecosystems, these enzymes are believed to play critical roles in P cycles (Speir and Ross 1978) as evidence shows that they are correlated to P stress and plant growth. Apart from being good indicators of soil fertility, phosphatase enzyme plays a key role in the soil system (Eivazi and Tabatabai 1977; Dick et al. 2000). For example, when there is a signal indicating P deficiency in the soil, acid phosphatase secretion from plant roots is increased to enhance the solubilization and remobilization of phosphate, thus influencing the ability of the plant to cope with P-stressed conditions (Karthikeyan et al. 2002; Mudge et al. 2002; Versaw and Harrison 2002). Understanding the dynamics of enzyme activities in these systems is crucial for predicting their interactions as their activities may, in turn, regulate nutrient uptake and plant growth, later on, where soil health is concerned.

2.5.8 Proteases

Proteases in the soil play a significant role in N mineralization (Ladd and Jackson 1982), an important process regulating the amount of plant available N and plant

growth. This enzyme in the soil is generally associated with inorganic and organic colloids (Burns 1982; Nannipieri et al. 1996). The amount of this extra cellular enzyme activity may be indicative not only of the biological capacity of soil for the enzymatic conversion of the substrate, which is independent of the extent of microbial activity, but might also have an important role in the ecology of microorganisms in the ecosystem (Burns 1982). There is a need to study the properties and factors affecting naturally occurring enzyme complexes such as those involving protease enzymes in the soil ecosystem as they may reveal some unknown role(s) in maintaining soil health and fertility.

2.5.9 Urease

Urease enzyme is responsible for the hydrolysis of urea fertilizers applied to the soil into NH_3 and CO_2 with the concomitant rise in soil pH (Andrews et al. 1989; Byrnes and Amberger 1989). This, in turn, results in a rapid N loss to the atmosphere through NH_3 volatilization (Simpson et al. 1984; Simpson and Freney 1988). Due to this role, urease activities in soils have received a lot of attention since it was first reported by Rotini (1935), a process considered vital in the regulation of N supply to plants after urea fertilization. Soil urease originates mainly from plants (Polacco 1977) and microorganisms found as both intra- and extra-cellular enzymes (Burns 1986; Mobley and Hausinger 1989). On the other hand, urease extracted from plants or microorganisms is rapidly degraded in soil by proteolytic enzymes (Pettit et al. 1976; Zantua and Bremner 1977). This suggests that a significant fraction of ureolytic activity in the soil is carried out by extracellular urease, which is stabilized by immobilization on organic and mineral soil colloids. Urease activity in soils is influenced by many factors. These include cropping history, organic matter content of the soil, soil depth, soil amendments, heavy metals, and environmental factors such as temperatures (Tabatabai 1977; Yang et al. 2006). For example, studies have shown that urease was very sensitive to toxic concentrations of heavy metals (Yang et al. 2006). Generally, urease activity increases with increasing temperature. It is suggested that higher temperatures increase the activity coefficient of this enzyme. Therefore, it is recommended that urea be applied at times of the day when temperatures are low. Since urease plays a vital role in the hydrolysis of urea fertilizer, it is important to uncover other unknown factors that may reduce the efficiency of this enzyme in the ecosystem.

2.6 Conclusion

It is very essential to understand the possible roles of soil enzymes in order to maintain soil health and its fertility management in ecosystems. These enzymes, usually found in the soil, may have significant effects on soil biology,

environmental management, growth and nutrient uptake in plants growing in ecosystems. Their activities may, however, be influenced by unknown cultural management practices either in a major or minor amount. Studies focusing the discovery of new enzymes from microbial diversity in the soil might be the most suitable practices that may positively influence their activities for improved plant growth as well as rendering the friendly biological environments in order to sustain other living beings.

References

- Acosta-Martínez V, Tabatabai MA (2000) Enzyme activities in a limed agricultural soil. *Biol Fertil Soils* 31:85–91
- Acton DF, Gregorich EG (1995) Executive summary. In: Acton DF, Gregorich EG (eds) *The health of our soils. Towards sustainable agriculture in Canada*. Center for Land and Biological Resources, Research Branch Agriculture and Agri-food Canada, ON, Canada
- Ajwa HA, Tabatabai MA (1994) Decomposition of different organic materials in soils. *Biol Fertil Soils* 18:175–182
- Alf K, Nannipieri P (1995) Cellulase activity, methods in applied soil microbiology and biochemistry. Academic, London
- Andrews RK, Blakeley RL, Zerner B (1989) Urease: a Ni (II) metalloenzyme. In: Lancaster JR (ed) *The bioinorganic chemistry of nickel*. VCH, New York, pp 141–166
- Arinze AE, Yubedee AG (2000) Effect of fungicides on *Fusarium* grain rot and enzyme production in maize (*Zea mays* L.). *Glob J Appl Sci* 6:629–634
- Atlas RM, Pramer D, Bartha R (1978) Assessment of pesticide effects on non-target soil microorganisms. *Soil Biol Biochem* 10:231–239
- Bandick AK, Dick RP (1999) Field management effects on soil enzyme activities. *Soil Biol Biochem* 31:1471–1479
- Baruah M, Mishra RR (1986) Effect of herbicides butachlor, 2,4-d and oxyfluorfen on enzyme activities and CO₂ evolution in submerged paddy field soil. *Plant Soil* 96:287–291
- Benckiser G, Santiago S, Neue HU, Watanabe I, Ottow JCG (1984) Effect of fertilization and exudation, dehydrogenase activity, iron-reducing populations and Fe²⁺ formation in the rhizosphere of rice (*Oryza sativa* L.) in relation to iron toxicity. *Plant Soil* 79:305–316
- Bloem J, de Ruiter P, Bouwman LA (1997) Food webs and nutrient cycling in agro-ecosystems. In: Van Elsas JD, Trevors JT, Wellington EMH (eds) *Modern soil microbiology*. Marcel Dekker, New York, pp 245–278
- Borner H (1958) Untersuchungen über den Abbau von Phlorizin im Boden. Ein Beitrag zum Problem der Bodenmudigkeit bei Obstgehölzen. *Naturwiss* 45:138–139
- Bromfield SM (1954) Reduction of ferric compounds by soil bacteria. *J Gen Microbiol* 11:1–6
- Brzezinska M, Stepniewska Z, Stepniewski W (1998) Soil oxygen status and dehydrogenase activity. *Soil Biol Biochem* 30:1783–1790
- Burns RG (1978) Enzyme activity in soil: some theoretical and practical considerations. In: Burns RG (ed) *Soil enzymes*. Academic, London, pp 295–340
- Burns RG (1982) Enzyme activity in soil: location and possible role in microbial ecology. *Soil Biol Biochem* 14:423–427
- Burns RG (1986) Interaction of enzymes with soil mineral and organic colloids. In: Huang PM, Schnitzer M (eds) *Interactions of soil minerals with natural organics and microbes*. Soil Science Society of America, Madison, pp 429–452
- Bynes BH, Amberger A (1989) Fate of broadcast urea in a flooded soil when treated with *N*-(*n*-butyl) thiophosphoric triamide, a urease inhibitor. *Fertil Res* 18:221–231

- Chet I (1987) Trichoderma-application, mode of action, and potential as biocontrol agent of soil borne pathogenic fungi. In: Chet I (ed) Innovative approaches to plant disease control. Wiley, New York, pp 137–349
- Chet I, Henis Y (1975) Sclerotial morphogenesis in fungi. *Annu Rev Phytopathol* 13:169–192
- Chet I, Ordentlich A, Shapira R, Oppenheim A (1990) Mechanism of biocontrol of soil borne plant pathogen by rhizobacteria. *Plant Soil* 129:85–92
- Deng SP, Tabatabai MA (1994) Cellulase activity of soils. *Soil Biol Biochem* 26:1347–1354
- Deshpande MV (1986) Enzymatic degradation of chitin and its biological applications. *J Sci Ind Res* 45:273–281
- Dick RP, Sandor JA, Eash NS (1994) Soil enzyme activities after 1500 years of terrace agriculture in the Colca Valley. *Peru Agric Ecosyst Environ* 50:123–131
- Dick RP, Breakwell DP, Turco RF (1996) Soil enzyme activities and biodiversity measurements as integrative microbiological indicators. In: Doran JW, Jones AJ (eds) Methods of assessing soil quality. Soil Science Society of America, Madison, WI, pp 247–271
- Dick RP (1997) Soil enzyme activities as integrative indicators of soil health. In: Pankhurst CE, Doube BM, Gupta VVSR (eds) Biological indicators of soil health. CABI, Wallingford, pp 121–156
- Dick WA, Cheng L, Wang P (2000) Soil acid and alkaline phosphatase activity as pH adjustment indicators. *Soil Biol Biochem* 32:1915–1919
- Dodgson KS, White G, Fitzgerald JW (1982) Sulphatase enzyme of microbial origin. *Afr J Biotechnol Vol I*. CRC, FL, pp 156–159
- Doran JW, Parkin TB (1994) Defining and assessing soil quality. In: Doran JW, Coleman DC, Bezdicek DF, Stewart BA (eds) Defining soil quality for a sustainable environment. Soil Science Society of America, Madison, pp 3–21
- Doran JW, Safley M (1997) Defining and assessing soil health and sustainable productivity. In: Pankhurst CE, Doube BM, Gupta VVSR (eds) Biological indicators of soil health. CABI, Wallingford, pp 1–28
- Eivazi F, Tabatabai MA (1977) Phosphates in soils. *Soil Biol Biochem* 9:167–172
- Eivazi F, Tabatabai MA (1988) Glucosidases and galactosidases in soils. *Soil Biol Biochem* 20:601–606
- Ellert BH, Clapperton MJ, Anderson DW (1997) An ecosystem perspective of soil quality. In: Gregorich EG, Carter MR (eds) Soil quality for crop production and ecosystem health. Elsevier, Amsterdam, pp 115–141
- Elliott LF, Lynch JM, Papendick RI (1996) The microbial component of soil quality. In: Stotzky G, Bollag JM (eds) Soil biochemistry. Marcel Dekker, New York, pp 1–21
- Eriksson KEL, Blanchette RA, Ander P (1990) Biodegradation of cellulose. In: Eriksson KEL, Blanchette RA, Ander P (eds) Microbial and enzymatic degradation of wood and wood components. Springer, New York, pp 89–180
- Esen A (1993) β -glucosidases: overview. In: Esen A (ed) β -glucosidases and molecular biology. American Chemical Society, Washington, DC, pp 9–17
- Frank T, Malkomes HP (1993) Influence of temperature on microbial activities and their reaction to the herbicide Goltix in different soils under laboratory conditions. *Zentralblatt für Mikrobiol* 148:403–412
- Galstian AS, Awungian ZS (1974) Significance of the enzymes in oxidation of Fe and Mn oxides in soil (in Russian). *Trans. 10th Intern. Congress Soil Sci III*. Nauka Publishing House, Moscow, pp 130–135
- Ganeshamurthy AM, Singh G, Singh NT (1995) Sulphur status and response of rice to sulphur on some soils of Andaman and Nicobar Islands. *J Indian Soc Soil Sci* 43:637–641
- Garcia C, Hernández T (1997) Biological and biochemical indicators in derelict soils subject to erosion. *Soil Biol Biochem* 29:171–177
- Glinski J, Stepniewski W (1985) Soil aeration and its role for plants. CRC, Boca Raton, FL
- Gupta VVSR, Farrell RE, Germida JJ (1993) Activity of arylsulphatases in Saskatchewan soils. *Can J Soil Sci* 73:341–347

- Halvorson JJ, Smith JL, Papendick RI (1997) Issues of scale for evaluating soil quality. *J Soil Water Conserv* 52:26–30
- James ES, Russel LW, Mitrick A (1991) Phosphate stress response in hydroponically grown maize. *Plant Soil* 132:85–90
- Kandeler E (1996) Nitrate. In: Schinner F, Öhlinger R, Kandeler E, Margesin R (eds) *Methods in soil biology*. Springer, Berlin, pp 408–410
- Kanfer JN, Mumford RA, Raghavan SS, Byrd J (1974) Purification of β -glucosidase activities from bovine spleen affinity chromatography. *Anal Biochem* 60:200–205
- Karthikeyan AS, Varadarajan DK, Mukatira UT, D'Urzo MP, Damaz B, Raghothama KG (2002) Regulated expression of *Arabidopsis* phosphate transporters. *Plant Physiol* 130:221–233
- Kennedy AC, Papendick RI (1995) Microbial characteristics of soil quality. *J soil water conserv.* May–June:243–248
- Kertesz MA, Mirleau P (2004) The role of soil microbes in plant sulphur nutrition. *J Exp Bot* 55:1939–1945
- King NJ (1967) Glucoamylase of *Coniophora cerebella*. *Biochem J* 105:577–583
- Kiss S, Dragan-Bularda M, Radulescu D (1978) Soil polysaccharidases: activity and agricultural importance. In: Burns RG (ed) *Soil enzymes*. Academic, London, pp 117–147
- Klose S, Tabatabai MA (1999) Arylsulphatase activity of microbial biomass in soils. *Soil Sci Soc Am J* 63:569–574
- Ladd JN, Jackson RB (1982) In: Stevenson FJ (ed) *Nitrogen in agricultural soils*. American Society of Agronomy, WI, pp 173–228
- Madejón E, Burgos P, López R, Cabrera F (2001) Soil enzymatic response to addition of heavy metals with organic residues. *Biol Fertil Soils* 34:144–150
- Martinez CE, Tabatabai MA (1997) Decomposition of biotechnology by-products in soils. *J Environ Qual* 26:625–632
- McCarthy GW, Siddaramappa R, Reight RJ, Coddling EE, Gao G (1994) Evaluation of coal combustion by products as soil liming materials: their influence on soil pH and enzyme activities. *Biol Fertil Soils* 17:167–172
- McGill WB, Colle CV (1981) Comparative aspects of cycling of organic C, N, S and P through soil organic matter. *Geoderma* 26:267–286
- McLaren AD (1975) Soil as a system of humus and clay immobilised enzymes. *Chem Scripta* 8:97–99
- Miwa T, Ceng CT, Fujisaki M, Toishi A (1937) Zur Frage der Spezifität der Glykosidasen. I. Verhalten von β -D-Glucosidasen verschiedener Herkunft gegenüber den β -D-Glucosiden mit verschiedenen Aglykonen. *Acta Phytochim (Tokyo)* 10:155–170
- Mobley HLT, Hausinger RP (1989) Microbial urease: significance, regulation and molecular characterization. *Microbiol Rev* 53:85–108
- Mudge SR, Rae AL, Diatloff E, Smith FW (2002) Expression analysis suggests novel roles for members of Pht1 family of phosphate transporters in *Arabidopsis*. *Plant J* 31:341–353
- Nannipieri P, Sequi P, Fusi P (1996) Humus and enzyme activity. In: Piccolo A (ed) *Humic substances in terrestrial ecosystems*. Elsevier, New York, pp 293–328
- Ndiaye EL, Sandeno JM, McGrath D, Dick RP (2000) Integrative biological indicators for detecting change in soil quality. *Am J Altern Agric* 15:26–36
- Ordentlich A, Elad Y, Chet I (1988) The role of chitinase of *Serratia marcescens* in biocontrol of *Sclerotium rolfsii*. *Phytopathology* 78:84–88
- Pancholy SK, Rice EL (1973) Soil enzymes in relation to old field succession; amylase, cellulose, invertase, dehydrogenase and urease. *Soil Sci Soc Am J* 37:47–50
- Pankhurst CE, Hawke BG, McDonald HJ, Kirkby CA, Buckerfield JC, Michelsen P, O'Brien KA, Gupta VVSR, Doube BM (1995) Evaluation of soil biological properties as potential bioindicators of soil health. *Aust J Exp Agric* 35:1015–1028
- Pankhurst CE, Doube BM, Gupta VVSR (1997) Biological indicators of soil health: synthesis. In: Pankhurst CE, Doube BM, Gupta VVSR (eds) *Biological indicators of soil health*. CABI, Wallingford, Oxfordshire, pp 419–435

- Park D (1960) Antagonism – the background of soil fungi. In: Parkinson D, Waid JS (eds) The ecology of soil fungi. Liverpool University Press, Liverpool, pp 148–159
- Parr JF, Papendick RI, Hornick SB, Meyer RE (1992) Soil quality: attributes and relationship to alternative and sustainable agriculture. *Am J Altern Agric* 7:5–11
- Patrick ZA (1955) The peach replant problem in Ontario. II. Toxic substances from microbial decomposition products of peach root residues. *Can J Bot* 33:461–486
- Pazur JH (1965) Enzymes in the synthesis and hydrolysis of starch. In: Whistler R, Paschall EF (eds) Starch: chemistry and technology, vol 1, Fundamental aspects. Academic, New York, pp 133–175
- Petker AS, Rai PK (1992) Effect of fungicides on activity, secretion of some extra cellular enzymes and growth of *Alternaria alternata*. *Indian J Appl Pure Biol* 7:57–59
- Pettit NM, Smith ARJ, Freedman RB, Burns RG (1976) Soil urease: activity, stability and kinetic properties. *Soil Biol Biochem* 8:479–484
- Pitchel JR, Hayes JM (1990) Influence of fly ash on soil microbial activity and populations. *J Environ Qual* 19:593–597
- Polacco JC (1977) Is nickel a universal component of plant ureases? *Plant Sci Lett* 10:249–255
- Reddy GB, Faza A (1989) Dehydrogenase activity in sludge amended soil. *Soil Biol Biochem* 21:327
- Richmond PA (1991) Occurrence and functions of native cellulose. In: Haigler CH, Weimer PJ (eds) Biosynthesis and biodegradation of cellulose. Marcel Dekker, New York, pp 5–23
- Ross DJ (1975) Studies on a climosequence of soils in tussock grasslands-5. Invertase and amylase activities of topsoils and their relationships with other properties. *NZ J Sci* 18:511–518
- Ross DJ (1976) Invertase and amylase activities in ryegrass and white clover plants and their relationships with activities in soils under pasture. *Soil Biol Biochem* 8:351–356
- Rotini OT (1935) La trasformazione enzimatica dell'urea nel terreno. *Ann Labor Ric Ferm Spallanrani* 3:143–154
- Sarathchandra SU, Perrott KW (1981) Determination of phosphatase and arylsulphatase activity in soils. *Soil Biol Biochem* 13:543–545
- Shapira R, Ordentlich A, Chet I, Oppenheim AB (1989) Control of plant diseases by chitinase expressed from cloned DNA in *Escherichia coli*. *Phytopathology* 79:1246–1249
- Shawale JG, Sadana J (1981) Purification, characterization and properties of β -glucosidase enzyme from *Sclerotium rolfisii*. *Arch Biochem Biophys* 207:185–196
- Simpson JR, Freney JR, Wetselaar R, Muirhead WA, Leuning R, Denmead OT (1984) Transformations and losses of urea nitrogen after application to flooded rice. *Aust J Agric Res* 35:189–200
- Simpson JR, Freney JR (1988) Interacting processes in gaseous nitrogen loss from urea applied to flooded rice fields. In: Pushparajah E, Husin A, Bachik AT (eds) Proceedings of international symposium on urea technology and utilization. Malaysian Society of Soil Science, Kuala Lumpur, pp 281–290
- Singh PP, Shin YC, Park CS, Chung YR (1999) Biological control of *Fusarium* wilt of cucumber by chitinolytic bacteria. *Phytopathology* 89:92–99
- Singer MJ, Ewing S (2000) Soil quality. In: Sumner ME (ed) Handbook of soil science. CRC, Boca Raton, FL, pp 271–298
- Sinsabaugh RL, Linkins AE (1989) Natural disturbance and the activity of *Trichoderma viride* cellulase complex. *Soil Biol Biochem* 21:835–839
- Sinsabaugh RL, Antibus RK, Linkins AE (1991) An enzymic approach to the analysis of microbial activity during plant litter decomposition. *Agric Ecosyst Environ* 34:43–54
- Speir TW, Ross DJ (1978) Soil phosphatase and sulphatase. In: Burns RG (ed) Soil enzymes. Academic, London, UK, pp 197–250
- Srinivasulu M, Rangaswamy V (2006) Activities of invertase and cellulase as influenced by the application of tridemorph and captan to groundnut (*Arachis hypogaea*) soil. *Afr J Biotechnol* 5:175–180

- Tabatabai MA (1977) Effect of trace elements on urease activity in soils. *Soil Biol Biochem* 9:9–13
- Tabatabai MA (1982) Soil enzyme. In: Page AL (ed) *Methods of soil analysis, Part 2*. American Society of Agronomy, Madison, WI, pp 903–948
- Tabatabai MA (1994a) Soil enzymes. In: Weaver RW, Angle JS, Bottomley PS (eds) *Methods of soil analysis, part 2. Microbiological and biochemical properties*. SSSA Book Series No. 5. Soil Science Society of America, Madison, WI, pp 775–833
- Tabatabai MA (1994b) Soil enzymes. In: Mickelson SH (ed) *Methods of soil analysis, Part 2. Microbiological and biochemical properties*. Soil Science Society of America, Madison, WI, pp 775–833
- Tate RL (1995) *Soil microbiology*. John Wiley, New York
- Thoma JA, Spradlin JE, Dygert S (1971) Plant and animal amylases. In: Boyer PD (ed) *The enzymes*, 5th edn. Academic, New York, pp 115–189
- Torstensson L, Pell M, Stenberg B (1998) Need of a strategy for evaluation of arable soil quality. *Ambio* 27:4–8
- Trevors JT (1984) Dehydrogenase activity in soil: a comparison between the INT and TTC assay. *Soil Biol Biochem* 16:673–674
- Tyler G (1981) Heavy metals in soil biology and biochemistry. In: Paul EA, Ladd JN (eds) *Soil biochemistry*, vol 5. Marcel Dekker, New York, pp 371–414
- Versaw WK, Harrison MJ (2002) A chloroplast phosphate transporter, PHT2; 1, influences allocation of phosphate within the plant and phosphate-starvation responses. *Plant Cell* 14:1751–1766
- Vincent PG, Sisler HD (1968) Mechanisms of antifungal action of 2,4,5,6-tetrachloroisophthalonitrile. *Physiol Plant* 21:1249–1264
- Vong PC, Dedouge O, Lasserre-Joulin F, Guckert A (2003) Immobilized-S, microbial biomass-S and soil arylsulphatase activity in the rhizosphere soil of rape and barley as affected by labile substrate C and N additions. *Soil Biol Biochem* 35:1651–1661
- White AR (1982) Visualization of cellulases and cellulose degradation. In: Brown RM (ed) *Cellulose and other natural polymer systems: biogenesis, structure, and degradation*. Plenum, New York, pp 489–509
- Wilke BM (1991) Effect of single and successive additions of cadmium, nickel and zinc on carbon dioxide evolution and dehydrogenase activity in a sandy Luvisol. *Biol Fertil Soils* 11:34–37
- Yang Z, Liu S, Zheng D, Feng S (2006) Effects of cadmium, zinc and lead on soil enzyme activities. *J Environ Sci* 18:1135–1141
- Zantua MI, Bremner JM (1977) Stability of urease in soils. *Soil Biol Biochem* 9:135–140



<http://www.springer.com/978-3-642-14224-6>

Soil Enzymology

Shukla, G.; Varma, A. (Eds.)

2011, XVI, 384 p. 75 illus., 6 illus. in color., Hardcover

ISBN: 978-3-642-14224-6