Preface

1. Building Plant Organs

1.1. Plant Organs: What Are They? Where Are They From? How Are They Connected?

Next to the clearly visible, above-ground parts—the leaves, flowers, fruits and stems—plants also comprise a less-visible half, hidden below ground: root systems. The green or colorful above-ground parts are essential for photosynthesis and reproduction (1, 2), while roots are important for nutrient and water uptake, anchoring, mechanical support, and storage (3–5). In contrast to mammals, plants generate new organs and tissues throughout their whole life. This often results in enormous organisms, such as giant sequoias or the Trembling Giant, a clonal colony of a single quaking aspen with a massive underground root system. Surprisingly, organs, such as leaves and lateral roots, are positioned at fairly regular spatial and temporal intervals and this requires tight coordination of the underlying molecular processes (3, 6). Especially since these organs develop from a subset of cells, often deeply embedded between various plant tissues. In addition, to control overall plant growth and reproduction, the various above- and below-ground organs need to communicate (7).

1.2. Model Systems

A good model is simple in structure, easy to study, to grow and to multiply, amenable to genetic analyses, and can increase our understanding of plant organogenesis fast. In the past decades, a lot of progress has been made by studying the model plant Arabidopsis thaliana or Thale Cress, and a recent special Plant Journal Issue was dedicated to this (8). However, for example, Arabidopsis has a small shoot apical meristem that is deeply buried between rosette leaves, is virtually impossible to access, and cannot be grown in culture. Thus, most studies on Arabidopsis organ initiation concern the induction of floral meristems from the inflorescence apex, which is more easily accessed (9–11). An alternative system is tomato because its vegetative shoot apical meristem is relatively large and therefore can be dissected without problems, grows vigorously under defined culture conditions, and is well suited for a wide variety of micromanipulations (6, 12, 13).

The first chapters of this book will give an up-to-date overview of the above- (Chapters 1 and 2) and below-ground parts (Chapters 3 and 4) in monocot and dicot plants. We especially highlight
aspects of why monocot and dicot roots are ideal model systems for organogenesis (Chapters 2 and 3). However, on the one hand we need to translate this information to economically interesting crops, or further investigate this directly in these crops (4, 14, 15). On the other hand, we need other simpler systems to understand the evolution of organs and to provide insight in the underlying molecular networks (e.g. Chapters 5 and 6). The chapters in this book provide exactly that, focusing on tools to study organogenesis in *Arabidopsis*, but also taking it further to cereal crops and highlighting emerging model systems.

While it is not always straightforward to translate particular techniques and approaches that work well in, for example, Arabidopsis to crop plants, several examples are discussed in this book on the level of shoot kinematics (Chapter 17), immunolocalization (Chapters 14 and 15), 3D root systems (Chapter 11) and a lateral root-inducible system (Chapter 9). In addition, to address specific questions, for example on the level of evolutionary biology, we need to start using other model systems. A few of these emerging model systems are introduced here, such as brown algae (Chapter 6), *Physcomitrella* (Chapter 2), and Podostemaceae (Chapter 5). More details on how to grow and study the brown alga *Ectocarpus* are provided in Chapter 22. There are obviously a number of other models for plant organogenesis that are not addressed here, but that have recently been reviewed, such as the *Arabidopsis* petal (16).

Organogenesis entails the regulation of cell division, cell expansion, cell- and tissue-type differentiation, and patterning of the organ as a whole. De novo organogenesis is especially important in plants, as most of plant development takes place post-embryonically. Therefore it is essential to gain insight into how organs are initiated and how they develop. However, this very often is subject to technical difficulties as these processes take place embedded deep in tissues or are difficult to access or visualize. Furthermore, plant cells are enclosed in a rigid wall making a tight control of the direction of polar cell growth and of the positioning of cell division planes very important for plant organogenesis. To study this, we need specialized techniques that are described in this book.

One of the very first steps in the development of a plant is the formation of ovules and embryos. The ovule and embryo of *Arabidopsis thaliana* have been established as an excellent model system with which to study organogenesis at the molecular and genetic level (17–19). How to study and image these structures is addressed in Chapters 9 and 18. A new plant organ develops from a subset of cells that has been specified, primed, etc. and which will undergo a series of cell divisions to give rise to a new plant part, such as a leaf, a flower, and a lateral root. To visualize the contribution of each cell and cell
division to building the mature organ, it is necessary to establish cell lineages. An elegant tool to achieve this is described in Chapter 13.

The totipotency of several plant cells is reflected in their ability to regenerate tissues and organs. An approach to study this is described in Chapter 21.

2. Novel Techniques

Due to the difficulties associated with studying particular processes, the development of novel, more sensitive techniques is essential. For example, the use of fluorescence-activated cell sorting (FACS) brought about a revolution in cell-specific analyses of transcriptomes and hormone levels in Arabidopsis (20, 21). Here, the use of this approach in the shoot apical meristem is described (Chapter 16). However, it is also important to get closer to the proteins, and as cell-specific proteome analyses are still difficult, other techniques have been developed. For example, ribosome pull down provides insight into the translatome (Chapter 19), and localizing RNAs and proteins in plants is useful (Chapters 20 and 21). In addition, classical genetics has its limitations, as exemplified through redundancy and embryo lethal mutations. To circumvent this, chemical genetics was put forward as an ideal tool, as described in Chapter 12.

3. Mathematical Modelling

Finally, as our knowledge increases, we need computer-based approaches to bring everything together. In several areas of plant organogenesis, this has been used successfully. Auxin has been a major focus of mathematical modelling, and this is reflected in a wide range of models describing the distribution and role of auxin (22–25). However, these in silico approaches are not always easy to use by wet-lab scientists. We therefore also need simpler, user-friendly systems, such as the one in Chapter 23.

References
